

WEST[Generate Collection](#)[Print](#)**Search Results - Record(s) 1 through 4 of 4 returned.**☐ 1. Document ID: US 6355438 B1

L1: Entry 1 of 4

File: USPT

Mar 12, 2002

US-PAT-NO: 6355438

DOCUMENT-IDENTIFIER: US 6355438 B1

TITLE: Method for quantitating oligonucleotides

DATE-ISSUED: March 12, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Baker; Brenda F.	Carlsbad	CA		
Yu; Zhengrong	Carlsbad	CA		
Leeds; Janet M.	Encinitas	CA		

US-CL-CURRENT: [435/6](#); [435/91.1](#), [435/91.2](#), [536/22.1](#), [536/23.1](#), [536/24.3](#), [536/24.33](#)

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequences	Attachments	Claims	KWIC
Draw Desc	Image										

☐ 2. Document ID: US 6348314 B1

L1: Entry 2 of 4

File: USPT

Feb 19, 2002

US-PAT-NO: 6348314

DOCUMENT-IDENTIFIER: US 6348314 B1

TITLE: Invasive cleavage of nucleic acids

DATE-ISSUED: February 19, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Prudent; James R.	Madison	WI		
Hall; Jeff G.	Madison	WI		
Lyamichev; Victor I.	Madison	WI		
Brow; Mary Ann D.	Madison	WI		
Dahlberg; James E.	Madison	WI		

US-CL-CURRENT: [435/6](#); [435/91.1](#)

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequences	Attachments	Claims	KWIC
Draw Desc	Image										

☐ 3. Document ID: US 6255081 B1

L1: Entry 3 of 4

File: USPT

Jul 3, 2001

US-PAT-NO: 6255081

DOCUMENT-IDENTIFIER: US 6255081 B1

TITLE: Thermostable flap endonuclease derived from hyperthermophile bacterium belonging to the genus pyrococcus

DATE-ISSUED: July 3, 2001

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Matsui; Ikuo	Ibaraki			JPX
Ishikawa; Kazuhiko	Ibaraki			JPX
Kosugi; Yoshitsugu	Ibaraki			JPX
Matsui; Eriko	Ibaraki			JPX
Kawasaki; Satoko	Chiba			JPX

US-CL-CURRENT: 435/91.1; 435/199, 435/252.3, 435/252.33, 435/320.1, 536/23.2

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequences	Attachments	Claims	RAWC
Draw Desc	Image										

☐ 4. Document ID: US 5874283 A

L1: Entry 4 of 4

File: USPT

Feb 23, 1999

US-PAT-NO: 5874283DOCUMENT-IDENTIFIER: US 5874283 A

TITLE: Mammalian flap-specific endonuclease

DATE-ISSUED: February 23, 1999

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Harrington; John Joseph	Shaker Heights	OH	44120	
Hsieh; Chih-Lin	St. Louis	MO	63131	
Lieber; Michael R.	St. Louis	MO	63131	

US-CL-CURRENT: 435/252.3; 435/199, 435/252.33, 435/320.1, 435/69.1, 530/350, 536/23.2, 536/23.5

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequences	Attachments	RAWC
Draw Desc	Image									

Generate Collection

Print

Terms	Documents
5874283	4

Display Format:

CIT

Change Format

Previous Page

Next Page

WEST

Generate Collection

Print

L1: Entry 2 of 4

File: USPT

Feb 19, 2002

US-PAT-NO: 6348314

DOCUMENT-IDENTIFIER: US 6348314 B1

TITLE: Invasive cleavage of nucleic acids

DATE-ISSUED: February 19, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Prudent; James R.	Madison	WI		
Hall; Jeff G.	Madison	WI		
Lyamichev; Victor I.	Madison	WI		
Brow; Mary Ann D.	Madison	WI		
Dahlberg; James E.	Madison	WI		

US-CL-CURRENT: 435/6; 435/91.1

CLAIMS:

We claim:

1. A method for detecting the presence of a target nucleic acid molecule by detecting non-target cleavage products comprising:

a) providing:

i) a cleavage agent;

ii) a source of target nucleic acid, said target nucleic acid comprising a first region and a second region, said second region downstream of and contiguous to said first region;

iii) a first oligonucleotide, wherein at least a portion of said first oligonucleotide is completely complementary to said first portion of said first target nucleic acid;

iv) a second oligonucleotide comprising a 3' portion and a 5' portion, wherein said 5' portion is completely complementary to said second portion of said target nucleic acid;

b) mixing said cleavage agent, said target nucleic acid, said first oligonucleotide and said second oligonucleotide to create a reaction mixture under reaction conditions such that at least said portion of said first oligonucleotide is annealed to said first region of said target nucleic acid and wherein at least said 5' portion of said second oligonucleotide is annealed to said second region of said target nucleic acid so as to create a cleavage structure, and wherein cleavage of said cleavage structure occurs to generate non-target cleavage product; and

c) detecting the cleavage of said cleavage structure.

2. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises detecting said non-target cleavage product.

3. The method of claim 1, wherein said 3' portion of said second oligonucleotide comprises a 3' terminal nucleotide not complementary to said target nucleic acid.

4. The method of claim 1, wherein said 3' portion of said second oligonucleotide consists of a single nucleotide not complementary to said target nucleic acid.
5. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises detection of fluorescence.
6. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises detection of mass.
7. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises detection of fluorescence energy transfer.
8. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises detection selected from the group consisting of detection of radioactivity, luminescence, phosphorescence, fluorescence polarization, and charge.
9. The method of claim 1, wherein said first oligonucleotide is attached to a solid support.
10. The method of claim 1, wherein said second oligonucleotide is attached to a solid support.
11. The method of claim 1, wherein said cleavage agent comprises a structure-specific nuclease.
12. The method of claim 11, wherein said structure-specific nuclease comprises a thermostable structure-specific nuclease.
13. The method of claim 11, wherein said cleavage agent comprises a 5' nuclease.
14. The method of claim 13, wherein said 5'-nuclease comprises a thermostable 5'-nuclease.
15. The method of claim 13, wherein a portion of the amino acid sequence of said nuclease is homologous to a portion of the amino acid sequence of a thermostable DNA polymerase derived from a thermophilic organism.
16. The method of claim 15, wherein said thermophilic organism is selected from the group consisting of *Thermus aquaticus*, *Thermus flavus*, and *Thermus thermophilus*.
17. The method of claim 1, wherein said target nucleic acid comprises DNA.
18. The method of claim 1, wherein said target nucleic acid comprises RNA.
19. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises:
 - a) providing:
 - i) said non-target cleavage product;
 - ii) a composition comprising two single-stranded nucleic acids annealed so as to define a single-stranded portion of a protein binding region; and
 - iii) a protein; and
 - b) exposing said non-target cleavage product to said single-stranded portion of said protein binding region under conditions such that said protein binds to said protein binding region.
20. The method of claim 19, wherein said protein comprises a nucleic acid producing protein and wherein said nucleic acid producing protein binds to said protein binding region and produces nucleic acid.
21. The method of claim 20, wherein said protein binding region is a template-dependent RNA polymerase binding region.
22. The method of claim 21, wherein said template-dependent RNA polymerase binding region is a T7 RNA polymerase binding region.

23. The method of claim 1, wherein said detecting the cleavage of said cleavage structure comprises:

a) providing:

i) said non-target cleavage product;

ii) a single continuous strand of nucleic acid comprising a sequence defining a single strand of an RNA polymerase binding region;

iii) a template-dependent DNA polymerase; and

iv) a template-dependent RNA polymerase;

b) exposing said non-target cleavage product to said RNA polymerase binding region under conditions such that said non-target cleavage product binds to a portion of said single strand of said RNA polymerase binding region to produce a bound non-target cleavage product;

c) exposing said bound non-target cleavage product to said template-dependent DNA polymerase under conditions such that a double-stranded RNA polymerase binding region is produced; and

d) exposing said double-stranded RNA polymerase binding region to said template-dependent RNA polymerase under conditions such that RNA transcripts are produced.

24. The method of claim 23, further comprising the step of e) detecting said RNA transcripts.

25. The method of claim 23, wherein said template-dependent RNA polymerase is T7 RNA polymerase.

26. The method of claim 1, wherein said target nucleic acid comprises single-stranded DNA.

27. The method of claim 1, wherein said target nucleic acid comprises double-stranded DNA and prior to step c), said reaction mixture is treated such that said double-stranded DNA is rendered substantially single-stranded.

28. The method of claim 27, wherein said double-stranded DNA is rendered substantially single-stranded by heat.

29. The method of claim 1, wherein said source of target nucleic acid comprises a sample containing genomic DNA.

30. The method of claim 29, wherein said sample is selected from the group comprising blood, saliva, cerebral spinal fluid, pleural fluid, milk, lymph, sputum and semen.

31. The method of claim 1, wherein said reaction conditions comprise providing a source of divalent cations.

32. The method of claim 31, wherein said divalent cation is selected from the group consisting of Mn.sup.2+ and Mg.sup.2+ ions.

33. The method of claim 1, wherein said first and said second oligonucleotides are provided in concentration excess compared to said target nucleic acid.

34. The method of claim 1, further comprising providing a third oligonucleotide complementary to a third portion of said target nucleic acid upstream of said first portion of said first target nucleic acid, wherein said third oligonucleotide is mixed with said reaction mixture in step b).

35. A method for detecting the presence of a target nucleic acid molecule by detecting non-target cleavage products comprising:

a) providing:

- i) a cleavage agent;
- ii) a source of target nucleic acid, said target nucleic acid comprising a first region and a second region, said second region downstream of and contiguous to said first region;
- iii) a plurality of first oligonucleotides, wherein at least a portion of said first oligonucleotides is completely complementary to said first portion of said first target nucleic acid;
- iv) a second oligonucleotide comprising a 3' portion and a 5' portion, wherein said 5' portion is completely complementary to said second portion of said target nucleic acid;
- b) mixing said cleavage agent, said target nucleic acid, said plurality of first oligonucleotides and said second oligonucleotide to create a reaction mixture under reaction conditions such that at least said portion of a first oligonucleotide is annealed to said first region of said target nucleic acid and wherein at least said 5' portion of said second oligonucleotide is annealed to said second region of said target nucleic acid so as to create a cleavage structure, and wherein cleavage of said cleavage structure occurs to generate non-target cleavage product, wherein said conditions permit multiple cleavage structures to form and be cleaved from said target nucleic acid; and
- c) detecting the cleavage of said cleavage structures.

36. The method of claim 35, wherein said conditions comprise isothermal conditions that permit said plurality of first oligonucleotides to dissociate from said target nucleic acid.

37. The method of claim 35, wherein said target nucleic forms a cleavage structure with two or more of said plurality of first oligonucleotides, wherein the cleavage structure is cleaved to produce said non-target cleavage products.

38. The method of claim 37, wherein said target nucleic forms a cleavage structure with ten or more of said plurality of first oligonucleotides, wherein the cleavage structure is cleaved to produce said non-target cleavage products.

39. The method of claim 37, wherein said target nucleic forms a cleavage structure with one hundred or more of said plurality of first oligonucleotides, wherein the cleavage structure is cleaved to produce said non-target cleavage products.

40. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises detecting said non-target cleavage product.

41. The method of claim 35, wherein said 3' portion of said second oligonucleotide comprises a 3' terminal nucleotide not complementary to said target nucleic acid.

42. The method of claim 35, wherein said 3' portion of said second oligonucleotide consists of a single nucleotide not complementary to said target nucleic acid.

43. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises detection of fluorescence.

44. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises detection of mass.

45. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises detection of fluorescence energy transfer.

46. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises detection selected from the group consisting of detection of radioactivity, luminescence, phosphorescence, fluorescence polarization, and charge.

47. The method of claim 35, wherein at least one of said plurality of first oligonucleotides is attached to a solid support.

48. The method of claim 35, wherein said second oligonucleotide is attached to a solid support.

49. The method of claim 35, wherein said cleavage agent comprises a structure-specific nuclease.
50. The method of claim 49, wherein said structure-specific nuclease comprises a thermostable structure-specific nuclease.
51. The method of claim 49, wherein said cleavage agent comprises a 5' nuclease.
52. The method of claim 51, wherein said 5'-nuclease comprises a thermostable 5'-nuclease.
53. The method of claim 51, wherein a portion of the amino acid sequence of said nuclease is homologous to a portion of the amino acid sequence of a thermostable DNA polymerase derived from a thermophilic organism.
54. The method of claim 53, wherein said thermophilic organism is selected from the group consisting of *Thermus aquaticus*, *Thermus flavus*, and *Thermus thermophilus*.
55. The method of claim 35, wherein said target nucleic acid comprises DNA.
56. The method of claim 35, wherein said target nucleic acid comprises RNA.
57. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises:
- a) providing:
 - i) said non-target cleavage product;
 - ii) a composition comprising two single-stranded nucleic acids annealed so as to define a single-stranded portion of a protein binding region; and
 - iii) a protein; and
 - b) exposing said non-target cleavage product to said single-stranded portion of said protein binding region under conditions such that said protein binds to said protein binding region.
58. The method of claim 57, wherein said protein comprises a nucleic acid producing protein and wherein said nucleic acid producing protein binds to said protein binding region and produces nucleic acid.
59. The method of claim 58, wherein said protein binding region is a template-dependent RNA polymerase binding region.
60. The method of claim 59, wherein said template-dependent RNA polymerase binding region is a T7 RNA polymerase binding region.
61. The method of claim 35, wherein said detecting the cleavage of said cleavage structures comprises:
- a) providing:
 - i) said non-target cleavage product;
 - ii) a single continuous strand of nucleic acid comprising a sequence defining a single strand of an RNA polymerase binding region;
 - iii) a template-dependent DNA polymerase; and
 - iv) a template-dependent RNA polymerase;
 - b) exposing said non-target cleavage product to said RNA polymerase binding region under conditions such that said non-target cleavage product binds to a portion of said single strand of said RNA polymerase binding region to produce a bound non-target cleavage product;
 - c) exposing said bound non-target cleavage product to said template-dependent DNA polymerase under conditions such that a double-stranded RNA polymerase binding region

is produced; and

d) exposing said double-stranded RNA polymerase binding region to said template-dependent RNA polymerase under conditions such that RNA transcripts are produced.

62. The method of claim 61, further comprising the step of e) detecting said RNA transcripts.

63. The method of claim 61, wherein said template-dependent RNA polymerase is T7 RNA polymerase.

64. The method of claim 35, wherein said target nucleic acid comprises single-stranded DNA.

65. The method of claim 35, wherein said target nucleic acid comprises double-stranded DNA and prior to step c), said reaction mixture is treated such that said double-stranded DNA is rendered substantially single-stranded.

66. The method of claim 65, wherein said double-stranded DNA is rendered substantially single-stranded by heat.

67. The method of claim 35, wherein said source of target nucleic acid comprises a sample containing genomic DNA.

68. The method of claim 67, wherein said sample is selected from the group comprising blood, saliva, cerebral spinal fluid, pleural fluid, milk, lymph, sputum and semen.

69. The method of claim 35, wherein said reaction conditions comprise providing a source of divalent cations.

70. The method of claim 69, wherein said divalent cation is selected from the group consisting of Mn.sup.2+ and Mg.sup.2+ ions.

71. The method of claim 35, wherein said plurality of first oligonucleotides and said second oligonucleotides are provided in concentration excess compared to said target nucleic acid.

72. The method of claim 35, further comprising providing a third oligonucleotide complementary to a third portion of said target nucleic acid upstream of said first portion of said first target nucleic acid, wherein said third oligonucleotide is mixed with said reaction mixture in step b).

WEST**End of Result Set**

Generate Collection

Print

L1: Entry 4 of 4

File: USPT

Feb 23, 1999

US-PAT-NO: 5874283

DOCUMENT-IDENTIFIER: US 5874283 A

TITLE: Mammalian flap-specific endonuclease

DATE-ISSUED: February 23, 1999

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Harrington; John Joseph	Shaker Heights	OH	44120	
Hsieh; Chih-Lin	St. Louis	MO	63131	
Lieber; Michael R.	St. Louis	MO	63131	

ASSIGNEE-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY	TYPE CODE
Harrington; John Joseph	Cleveland	OH			04
Lieber; Michael R.	Arcadia	CA			04
Hsieh; Chih-Lin	Arcadia	CA			04

APPL-NO: 8/ 455968 [PALM]

DATE FILED: May 30, 1995

INT-CL: [6] C12 N 1/20, C12 N 9/22, C07 H 21/04, C07 K 1/00

US-CL-ISSUED: 435/252.3; 435/199, 435/252.33, 435/320.1, 435/69.1, 536/23.2, 536/23.5, 530/350

US-CL-CURRENT: 435/252.3; 435/199, 435/252.33, 435/320.1, 435/69.1, 530/350, 536/23.2, 536/23.5

FIELD-OF-SEARCH: 435/252.3, 435/199, 435/69.1, 435/252.33, 435/320.1, 536/23.2, 536/23.5, 530/350

PRIOR-ART-DISCLOSED:

U.S. PATENT DOCUMENTS

Search Selected

Search ALL

	PAT-NO	ISSUE-DATE	PATENTEE-NAME	US-CL
<input type="checkbox"/>	<u>5324830</u>	June 1994	Resnick et al.	536/23.2
<input type="checkbox"/>	<u>5359047</u>	October 1994	Donahue et al.	536/23.5

OTHER PUBLICATIONS

Chow and Fraser, "Purification and Properties of Single Strand DNA-binding Endo-Exonuclease of Neurospora crassa, " J. Biol. Chem., 258:12010-12018 (1983).
Chow and Resnick, "Purification and Characterization of an Endo-exonuclease from

Saccharomyces cerevisiae That is Influenced by the RAD52 Gene," J. Biol. Chem., 262:17659-17667 (1987).
Chow and Resnick, "An endo-exonuclease activity of yeast that requires a functional RAD52 gene," Molec. Gen. Genet., 211:41-48 (1988).
Habraken et al., "Yeast excision repair gene RAD2 encodes a single-stranded DNA endonuclease," Nature, 366:365-368 (1993).
Harosh et al., "Purification and Characterization of Rad3 ATPase/DNA Helicase from Saccharomyces cerevisiae," J. Biol. Chem., 264:20532-20539 (1989).
Harrington and Lieber, "Functional domains within FEN-1 and RAD2 define a family of structure-specific endonucleases: implications for nucleotide excision repair," Genes and Devel., 8:1344-1355 (1994).
Harrington and Lieber, "The characterization of a mammalian DNA structure-specific endonuclease," EMBO Journal, 13:1235-1246 (1994).
Harrington and Lieber, "DNA Structural Elements Required for FEN-1 Binding," J. Biol. Chem., 270:1-6 (1995).
Hiraoka et al., "Sequence of Human FEN-1, a Structure-Specific Endonuclease, and Chromosomal Localization of the gene (FEN1) in Mouse and Human," Genomics, 25:220-225 (1995).
Koa et al., "Endo-exonuclease of Aspergillus nidulans," Biochem. Cell Biol, 68:387-392 (1990).
Lyamichev et al., "Structure-Specific Endonucleolytic Cleavage of Nucleic Acids by Eubacterial DNA Polymerases," Science, 260:778-783 (1993).
Park et al., "RAD25 (SSL2), the yeast homolog of the human xeroderma pigmentosum group B DNA repair gene, is essential for viability," Proc. Natl. Acad. Sci., 89:11416-11420 (1992).
Sung et al., "RAD3 protein of Saccharomyces cerevisiae is a DNA helicase," Proc. Natl. Acad. Sci., 84:8951-8955 (1987).
Szankasi and Smith, "A Role for Exonuclease I from S. pombe in Mutation Avoidance and Mismatch Correction," Science, 267:1166-1168 (1995).
Tomkinson et al., "Yeast DNA repair and recombinant proteins Rad1 and Rad10 constitute a single-stranded-DNA endonuclease," Nature, 362:860-862 (1993).
Hiroaka et al. Genomics 25(1): 220-5, Jan. 1, 1995.
Murray et al. Molecular and Cellular Biology 14(7): 4878-88, Jul. 1994.

ART-UNIT: 162

PRIMARY-EXAMINER: Wax; Robert A.

ASSISTANT-EXAMINER: Saidha; Tekchand

ATTY-AGENT-FIRM: Townsend & Townsend & Crew

ABSTRACT:

Compositions comprising human FEN-1(flapp) endonucleases, nucleic acids encoding them, and methods for their use are provided.

6 Claims, 12 Drawing figures